

## ENDOFAUNA OF HELMINTH PARASITES OF FISH IN THE AMAZONIC BASIN

## ENDOFAUNA DE HELMINTOS PARASITOS DE PEIXES DA BACIA AMAZÔNICA

Lucena Rocha Virgilio<sup>1\*</sup>, Fabricia da Silva Lima<sup>2</sup>, Ricardo Massato Takemoto<sup>3</sup>, Luís Marcelo Aranha Camargo<sup>4</sup>, Dionatas Ulises de Oliveira Meneguetti<sup>5</sup>

1. Programa de Pós-Graduação em Biodiversidade e Biotecnologia (Bionorte), Universidade Federal do Acre (UFAC), Rio Branco, Acre, Brasil.
2. Laboratório de ecologia aquática, Universidade Federal do Acre (UFAC), Cruzeiro do Sul, *Campus* Floresta, Acre, Brazil.
3. Núcleo de Pesquisas em Limnologia, Ictiologia e Aquicultura, Laboratório de Ictioparasitologia, Universidade Estadual de Maringá, Maringá, Paraná, Brasil.
4. Instituto de Ciências Biomédicas da Universidade de São Paulo (ICB5 USP), Monte Negro, Rondônia, Brasil.
5. Laboratório de Medicina Tropical, Universidade Federal do Acre, Programa de Pós-Graduação Stricto Sensu em Ciências da Saúde na Amazônia Ocidental, Rio Branco, Acre, Brasil.

\* Autor correspondente: e-mail: [lurubita@gmail.com](mailto:lurubita@gmail.com)

### RESUMO

A biodiversidade de parasitos na Amazônia é subestimada em função da grande diversidade de peixes e elevado grau de endemismo, que favorecem a megadiversidade da fauna helmíntica na região. Assim, o presente estudo investigou a endofauna de helmintos de 13 espécies de peixes do alto Rio Juruá, estado do Acre, no sudoeste da Amazônia, Brasil. Os peixes foram capturados nos rios Juruá, Crôa, Paranã e Môa e os helmintos analisados conforme literatura especializada. Foram encontrados um total de 919 helmintos pertencentes a 25 espécies, sendo 9 espécies de Nematoda, 8 Cestoda, 6 Digenea e 2 Acanthocephalus. Entre as espécies de helmintos, *Procamallanus (S.) inopinatus* foi o mais predominante e apresentou maior intensidade de infestação e abundância. Novos relatos de peixes hospedeiros de helmintos foram verificados para *Cichla nigromaculata* parasitado por *Procamallanus (S.) inopinatus*, *Chaetobranchius favescens* hospedeiro de *Cosmoxynemoides agurei*, *Curimatella meyeri* para *Cosmoxynema vianai*, *Pseudoplatystoma fasciatum* para *Goezeella* sp., toda a endofauna de *Heros severus*, *Leporinus jamesi* para *Gibsoniela* sp., *Myloplus rubripinnis* para Digenea *Auriculostoma* sp. e *Opsodoras morei* para o Acanthocephalus *Sharpiloseptis peruviansis*. Assim, o presente estudo contribuiu com informações que aumentam registro desses parasitos na Amazônia brasileira, sugerindo que essa área apresenta um grande potencial para futuros estudos.

**Palavras-chaves:** Peixe; Amazônia; Helmintofauna; Acre.

### ABSTRACT

The Amazonian parasite biodiversity is underestimated due to the high fish diversity and a high degree of endemism, which favor the megadiversity of helminth fauna in the region. Thus, the present study investigated the helminth endofauna of 13 fish species from the upper Juruá River, state of Acre, in southwestern Amazon, Brazil. The fish were caught in the Juruá, Crôa, Paranã, and Môa rivers and the helminths were analyzed according to specialized literature. We found a total of 919 helminths distributed in 25 species, with nine species of Nematoda, eight of Cestoda, six of Digenea, and two Acanthocephalus. Among the helminth species, *Procamallanus (S.) inopinatus* was the most predominant and had the highest intensity of infestation and abundance. New records of fish hosting helminths were found for *Cichla nigromaculata* parasitized by *Procamallanus (S.) inopinatus*, *Chaetobranchius favescens* hosting *Cosmoxynemoides agurei*, *Curimatella meyeri* hosting *Cosmoxynema vianai*, *Pseudoplatystoma fasciatum* hosting *Goezeella* sp., all endofauna of *Heros severus*, *Leporinus jamesi* hosting *Gibsoniela* sp., *Myloplus rubripinnis* hosting Digenea *Auriculostoma* sp., and *Opsodoras morei* hosting Acanthocephalus *Sharpiloseptis peruviansis*. Thus, the present study increased the records of these parasites in the Brazilian Amazon, suggesting that this area has high potential for future studies.

**Keywords:** Fish; Amazon; Helminth fauna; Acre.

## 1. INTRODUCTION

The diversity of parasites in fish in the Amazon is underestimated due to the highly diverse ichthyofauna and high degree of endemism, which favors the megadiversity of helminth fauna in the region [1].

The fish-parasite relationship has been the subject of many studies that have addressed the roles of interactive and non-interactive processes since parasites are attractive models to studies on the organization and structure of parasitic communities in wild fish populations [2],[3]. Parasites are considered essential and integral elements in aquatic ecosystems where they conduct fundamental ecological processes. For example, they can influence the productivity and structure of an ecosystem's food chain [4],[5]. A healthy ecosystem, i.e., functional [6] is, therefore, a system rich in parasite species [7]. Thus, knowledge regarding the diversity of fish parasites can be a useful tool for the conservation of global biodiversity [1].

However, despite the great diversity of helminth species recorded in the Amazon, knowledge about the parasite richness remains incomplete in some places. Most studies are concentrated in Central and Eastern Amazon [8],[9],[10], with few studies in southwestern Amazon. Most authors have studied the distribution, composition, and influence of the parasite-host relationship in fish from farming systems and few in natural environments [11],[12],[13]. Besides, studies in the Juruá River basin have shown only parasitic fauna in cultivation systems [14]. In other words, there is a huge gap in studies regarding the biodiversity of parasites associated with fish in natural systems in this region of southwestern Amazon.

However, fish from natural environments usually harbor several taxa of endoparasites with different strategies and life cycles [15]. It is estimated that more than 10,000 species, belonging to many phyla, parasitize fish, with constant descriptions of new species [16]. Thus, the present study assumes that these poorly studied regions may present high parasite richness, providing additional information on species distribution and occurrence.

Therefore, the present study aimed to describe the helminth fauna of 13 fish species of the Juruá River basin in the southwestern Amazon, Brazil.

## 2. MATERIAL AND METHODS

The study was carried out in the municipality of Cruzeiro do Sul, State of Acre, Brazil (07°37'52" S and 72°40'12" W). The fish were captured with authorization by the Brazilian Institute of Environment and Renewable Natural Resources (No. 59642-2/2019) in the Juruá, Crôa, Paranã, and Môa rivers (Fig. 1), from January to October 2019. The captured fish (Table

1) were sent to the Laboratory of Aquatic Ecology, where they were identified [17],[18] measured, and weighed. Each fish was submitted to necropsy, and the gastrointestinal tract and viscera were examined using a stereomicroscope and microscope with 40 to 100x magnification.

The collection, fixation, preservation, and preparation of the parasites for identification followed the recommendations of [19]. The parasite species were identified according to Schmidt [20], [21], [22], [23], [24], [25]. The ecological terms used were adopted from those recommended by [26].

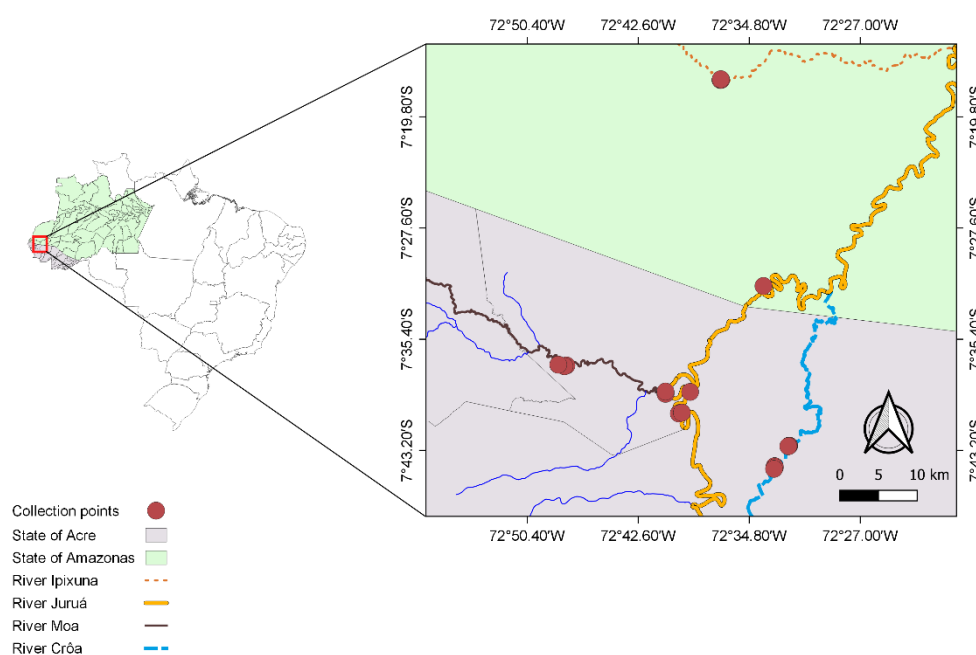


Fig. 1. Location of endoparasite collection points in southwestern Amazon, Acre, Brazil, 2019.

### 3. RESULTS

We collected 389 individuals of 13 fish species hosting a total of 919 helminths belonging to 25 species, with nine species of Nematoda, eight of Cestoda, six of Digenea, and two of Acanthocephala (Table 1).

Among the helminth species collected, *Procamallanus (Spirocamallnus) inopinatus* was found in the largest number of hosts, where it showed the highest intensity of infection (MI = 211.5) and mean abundance (MA = 105.8) in *Myloplus rubripinnis* (Table 1). Most hosts had adult Nematodes, and the only species in the larval stage in this group was *Contracaecum* sp. (larvae), which was found parasitizing *Pellona castelnaeana* (Table 1).

*Pseudoplatystoma fasciatum* was the host with the highest number of endoparasite species (N = 7), where six helminths belonged to the Cestoda group (Tables 1 and 2).

*Prochilodus nigricans*, *Pseudoplatystoma fasciatum*, *Hoplias malabaricus*, and *Serrasalmus* sp. had the highest parasite prevalence (~60%), while *Heros severus* had the lowest prevalence (16.6%) of endoparasites (Table 2).

In this study, we report new host-parasite relationships, such as *Chaetobranchus flavescens* parasitized by *Cosmoxynemoides aguirrei* (Figure 2A); *Curimatella meyeri* parasitized by *Cosmoxynema vianai* (Figure 2B); *Cichla nigromaculata* parasitized by *Procamallanus (Spirocamallanus) inopinatus* (Figure 2C); *Pseudoplatystoma fasciatum* parasitized by *Goezeella* sp. (Figure 3A); all endofauna of *Heros severus* and *Leporinus jamesi* parasitized by *Gibsoniela* sp. (Figure 3B); *Opsodoras morei* parasitized by *Sharpilosentis peruviensis* (Figure 3C); *Myloplus rubripinnis* parasitized by *Auriculostoma* sp. (Fig. 3D) (Table 1).



Fig.2 Nematodes reported in new host fish from rivers of the southwestern Amazon, Acre, Brazil, 2019. A- *Cosmoxynemoides aguirrei*, male; B- *Cosmoxynema vianai*, female; C- *Procamallanus (Spirocamallanus) inopinatus*, female. 1- Anterior part; 2- Posterior part.



Fig.3 A- *Goezeella* sp.; B- *Gibsoniela* sp.; C- *Sharpilosentis peruviensis*; D- *Auriculostoma* sp. collected in fish from the upper Juruá River, a tributary of the Amazon River, northern Brazil, Acre, 2019.

Table 1. Helminth species found in 13 hosts from the upper Juruá River, a tributary of the Amazon River, Acre, northern Brazil, 2019. P: Prevalence, MI: Mean intensity of infection, MA: Mean abundance, TNP: Total number of parasites. Nem.: Nematoda, Dig.: Digenea, Cest.: Cestoda, Acant.: Acanthocephala,

Host species	Parasite species	P (%)	MI	MA	Group	TNP
<i>Procamallanus</i>						
<i>Cichla nigromaculata</i> (Spirocamallanus)						
Kullander & Ferreira, 2006	<i>inopinatus</i> Travassos, Artigas & Pereira, 1928.	54.5	4.3	2.4	Nem.	26
<i>Chaetobranchus favescens</i> Heckel, 1840	<i>Cosmoxynemoides aguirrei</i> Travassos, 1949	15.8	1.6	0.4	Nem.	37
<i>Curimatella meyeri</i> Steindachner, 1882	<i>Cosmoxynema vianai</i> Travassos, 1949	40.0	1.8	0.7	Nem.	29

	Cladorchiidae		25.0	1.8	0.3	Dig.	1
<i>Pseudoplatystoma</i>							
<i>fasciatum</i>	Linnaeus, <i>Choanoscolex abscissus</i>						
1766	Riggenbach, 1896		100.0	16.0	16.0	Cest.	48
	<i>Nomimoscolex sudobim</i>						
	Woodland, 1935		66.7	2.7	4.0	Cest.	8
	<i>Goezeella</i> sp.		66.7	4.3	6.5	Cest.	13
	Proteocephalidae		66.7	4.0	6.0	Cest.	12
	<i>Dichelyne moraveci</i> Petter,						
	1995		66.7	2.7	4.0	Nem.	8
	<i>Spasskyellina spinulifera</i>						
	Woodland, 1935		33.3	2.7	8.0	Cest.	8
	<i>Monticellia</i> sp.		33.3	1.7	5.0	Cest.	5
<i>Acestrorhynchus</i>	<i>Prosthenhystera obesa</i>						
<i>falcatus</i> Bloch, 1794	Diesing, 1850		42.9	1.7	0.7	Dig.	10
	Pseudophyllidea		12.5	1.0	0.1	Cest.	1
	<i>Procamallanus</i>						
	( <i>Spirocamallanus</i> )						
<i>Heros severus.</i>	<i>inopinatus</i>		12.5	1.0	0.1	Nem.	1
	<i>Clinostomum</i> sp. (larvae)		25.0	2.0	0.5	Dig.	4
	<i>Ithyoclinostomum</i>						
	<i>dimorphum</i> Diesing, 1850						
	(larvae)		12.5	1.0	0.1	Dig.	1
<i>Prochilodus nigricans</i>	<i>Neoechinorhynchus</i>						
Agassiz, 1829	<i>curemai</i> Noronha, 1973		75.3	2.9	2.2	Acant.	178
<i>Leporinus jamesi</i>							
Garman, 1929	<i>Gibsoniela</i> sp.		33.3	6.0	2.0	Cest.	6
	<i>Deltamphistoma</i>						
<i>Myloplus rubripinnis</i>	<i>pitingaense</i> Thatcher &						
Müller & Troschel, 1844	Jégu, 1996		25.0	35.0	8.8	Dig.	35
	<i>Procamallanus</i>						
	( <i>Spirocamallanus</i> )						
	<i>inopinatus</i>		50.0	211.5	105.8	Nem.	423

	<i>Auriculostoma</i> sp.	100.0	1.0	0.3	Dig.	1
	<i>Sharpilosentis peruviansis</i>					
<i>Opsodoras morei</i>	Lisitsyna, Scholz & Steindachner, 1881	50.0	2.4	4.8	Acant.	19
<i>Pellona castelnaeana</i>						
Valenciennes, 1847	<i>Contraecum</i> sp. (larvae)	41.7	4.8	2.0	Nem.	24
<i>Hoplias malabaricus</i>	<i>Paraseuratum soaresi</i>					
Bloch, 1794	Fábio, 1983	66.7	1.5	1.0	Nem.	12
<i>Serrasalmus</i> sp.	<i>Raphidascaris</i> sp.	84.6	1.4	1.15	Nem.	15

Table 2. Host fish community of helminths from the upper Juruá River, a tributary of the Amazon River, Acre, northern Brazil, 2019.

Host species	Examined		P (%)	Total parasites
	(N)	Parasitized (N)		
<i>Cichla nigromaculata</i>	11	6	54.5	26
<i>Chaetobranchius favescens</i>	145	23	15.86	37
<i>Curimatella meyeri</i>	34	13	38.2	24
<i>Pseudoplatystoma fasciatum</i>	3	3	100	102
<i>Acestrorhynchus falcatus</i>	26	7	26.9	11
<i>Heros severus</i>	24	4	16.6	6
<i>Prochilodus nigricans</i>	81	61	75.3	178
<i>Leporinus jamesi</i>	3	1	33.3	6
<i>Myloplus rubripinnis</i>	9	4	44.4	459
<i>Opsodoras morei</i>	16	8	50	19
<i>Pellona</i> sp.	12	5	41.6	24
<i>Hoplias malabaricus</i>	12	8	66.6	12
<i>Serrasalmus</i> sp.	13	11	84.6	15

#### 4. DISCUSSION

The helminth species of the Nematoda and Cestoda groups predominated in this study, where Nematoda occurred in a greater variety of hosts. Generally, nematodes are more prevalent in neotropical fish assemblages in different ecosystems [27],[28].

Nematodes such as *Paraseuratum soaresi*, found in *Hoplias malabaricus*, *Raphidascari* sp. in *Serrasalmu* sp., and *Dichelyne moraveci* in *Pseudoplatystoma fasciatum* showed a high prevalence in those host species. This fact may be associated with the carnivorous habit of those fish [29], which consume organisms that are part of the life cycle of those helminths. According to [30], the diet of fish has a strong relationship with the incidence of endoparasite species since they include intermediate, parathenic, and definitive hosts in the environment, supporting the life cycle of those nematodes.

In the present study, most fish species studied were parasitized by adult nematodes, indicating that these hosts may be occupying a final position in the food web. [22] showed that this group of helminths has indirect life cycles and that fish can act as final and intermediate hosts.

The most common Nematoda was *Procamallanus (S.) inopinatus*, with a high degree of infestation in its hosts. This helminth is very generalist, occurring in many Amazonian freshwater fish species, which probably act as post-cyclical or para-definitive hosts [22].

The only Nematoda in the larval phase (L3) observed in the present study was *Contracaecum* sp. parasitizing *Pellona castelnaeana*. Many fish species can act as intermediate and/or parathenic hosts for larvae of *Contracaecum* sp., demonstrating a lack of specificity regarding the intermediate host [22],[31]. Thus, the present study indicated yet another report of these helminth larvae hosting fish. Since *P. castelnaeana* is highly appreciated by the riverside population and it is commercialized in local markets in the Amazon region [32], we suggest careful consumption due to the presence of these larvae with zoonotic potential.

Of the eight species of Cestoda found, six were in the intestine of *P. fasciatum*. These helminth species parasitizing *Pseudoplatystoma* have been studied in recent years [33],[34],[35]. The coexistence of those species in the intestine of a single host is common. Studies (including this study) have recorded the occurrence of Cestoda such as *Choanoscolex abscissus*, *Spasskyelina spinulifera*, *Nomimoscolex sudobim*, and *Monticellia* sp., in a single fish [36],[37],[38]. However, no study had recorded the occurrence of *Goezeella* sp. interacting with other Cestodes in the intestine of *P. fasciatum*.

*Gibsoniela* sp. was reported for the first time parasitizing *Leporinus jamesi*. Helminths of this genus have hitherto been found parasitizing only species of the genus *Ageneiosus*



(Siluriformes), such as *Gibsoniela mandube* and *Gibsoniela meursaulti* [39],[40]. This predominance of Proteocefalidae species evidenced in this study can also be associated with the carnivorous and omnivorous eating habits of those fish species, considering that most intermediate hosts of Cestoda are found in the diet of those species [41],[42].

Digenea was the third group with the highest prevalence in fish species, with the first *metacercariae* of *Clinostomum* sp. and *Ithyoclinostomum dimorphum* in the intestine of *Heros severus*. *Clinostomum* sp. and *Ithyoclinostomum dimorphum*, both from the family Clinostomidae and the first being a cosmopolitan genus [43],[44], are found in the adult stage in the esophagus of birds [45],[46]. They also parasitize many species of fish and mollusks that can be considered their intermediate hosts [47],[48].

*Auriculostoma* sp. (Digenea) was also the first report for the host species *Myloplus rubripinnis* in the Brazilian Amazon. In Brazil, this genus of helminths was found only in the south and southwest regions [49],[50],[51]. The *Auriculostoma* species closest to the Brazilian Amazon basin have been described in snails in the Peruvian Amazon, with *Auriculostoma foliaceum* described in *Bryconops caudomaculatus* and *Auriculostoma diagonale* in *Stethaprion erythropros* [52].

Only two fish species showed infestation by *Acanthocephalus*, with *Prochilodus nigricans* being parasitized by *Neochinorhynchus curemai* and *Opsodoras morei* by *Sharpilosentis peruviensis*. Characiform fish are the most common hosts for *Acanthocephalus* in the neotropical region, while few species have been reported in Siluriformes [53]. *Neochinorhynchus curemai* has been reported to infest *Prochilodus* species [54],[55] which was first described parasitizing *P. nigricans* in an Amazonian floodplain [56]. *Sharpilosentis peruviensis* (*Acanthocephala*) has been reported in Siluriformes from the Peruvian Amazon, being found in *Oxydoras niger* of the Doradidae family [57]. In the present study, this parasite is mentioned for the first time in Brazil, in *Opsodoras morei*, also from the Doradidae family, expanding the number of hosts and known distribution of this species.

## 5. CONCLUSION

The present study indicates new reports of eight fish species hosting endoparasites in new locations. Besides, it increases the records of these parasites for the Brazilian Amazon, such as *Sharpilosentis peruviensis* (*Acanthocephala*) and *Auriculostoma* sp. (Digenea), suggesting that this area has a high potential for future studies.

## 6. REFERENCES

- [1] LUQUE, J.L.; PEREIRA, F.B.; ALVES, P.V.; OLIVA, M.E.; TIMI, J.T. Helminth parasites of South American fishes: current status and characterization as a model for studies of biodiversity. **Journal of helminthology**. 91: 150-164, 2017. <https://doi.org/10.1017/S0022149X16000717>
- [2] HOSHINO, M.D.F.G.; NEVES, L.R.; TAVARES-DIAS, M. Parasite communities of the predatory fish, *Acestrorhynchus falcatus* and *Acestrorhynchus falcistrostris*, living in sympatry in Brazilian Amazon. **Revista Brasileira de Parasitologia Veterinária** 25:207-216, 2016. <https://doi.org/10.1590/S1984-29612016038>
- [3] RUEHLE, B.P.; HERRMANN, K.K.; HIGGINS, C.L. Helminth parasite assemblages in two cyprinids with different life history strategies. **Aquatic Ecology** 51:247-256, 2017. <https://doi.org/10.1007/s10452-017-9614-7>
- [4] POULIN, R. A importância funcional dos parasitas nas comunidades animais: muitos papéis em vários níveis ?. **Jornal internacional de parasitologia** , v. 29, n. 6, pág. 903-914, 1999.
- [5] MARCOGLIESE, D.J. Parasites of the superorganism: are they indicators of ecosystem health. **International journal for parasitology** 35:705-716, 2005. <https://doi.org/10.1016/j.ijpara.2005.01.015>
- [6] COSTANZA, R.; MAGEAU, M. What is a healthy ecosystem?. **Aquatic ecology**, v. 33, n. 1, p. 105-115, 1999.
- [7] HUDSON, P. J.; DOBSON, A. P.; LAFFERTY, K. D. Ecossistema saudável é rico em parasitas? **Tendências em ecologia e evolução**, v. 21, n. 7, pág. 381-385, 2006.
- [8] BAIA, R. R. J.; FLORENTINO, A. C.; SILVA, L. M. A.; & TAVARES-DIAS, M. Patterns of the parasite communities in a fish assemblage of a river in the Brazilian Amazon region. **Acta parasitologica**, 63(2), 304-316, 2018.
- [9] CORRÊA, L. L.; OLIVEIRA, M. S. B.; TAVARES-DIAS, M. Helminthic endofauna of four species of fish from lower Jari river, a tributary of the Amazon basin in Brazil. **Boletim do Instituto de Pesca**, v. 45, n. 1, 2019.
- [10] MORAIS, A. M.; CÁRDENAS, M. Q.; MALTA, J. C. O. Nematofauna of red piranha *Pygocentrus nattereri* (Kner, 1958) (Characiformes: Serrasalminidae) from Amazonia, Brazil. **Revista Brasileira de Parasitologia Veterinária**, v. 28, n. 3, p. 458-464, 2019.
- [11] CAVALCANTE, P.H.; SILVA, M.T.; SANTOS, E.G.; CHAGAS-MOUTINHO, V.A.; SANTOS, C.P. *Orientatractis moraveci* n. sp. and *Rondonia rondoni* Travassos, 1920 (Nematoda: Atractidae), parasites of *Pimelodus blochii* (Osteichthyes, Pimelodidae) from the

- Acre and Xapuri Rivers, Western Amazon, Brazil. *Parasitology* 144: 226-236, 2017. <https://doi.org/10.1017/S0031182016001736>
- [12] NEGREIROS, L.P.; PEREIRA, F.B.; TAVARES-DIAS, M.; TAVARES, L.E. Community structure of metazoan parasites from *Pimelodus blochii* in two rivers of the western Brazilian Amazon: same seasonal traits, but different anthropogenic impacts. *Parasitology research* 117: 3791-3798, 2018. <https://doi.org/10.1007/s00436-018-6082-5>
- [13] NEGREIROS, L.P.; PEREIRA, F.B.; TAVARES-DIAS, M. *Dadaytrema oxycephala* (Digenea: Cladorchiidae) in definitive host *Pimelodus blochii* (Pisces: Pimelodidae), with morphological and geographic distribution data in fishes from the South America. *Journal of Parasitic Diseases* 44:62-68, 2020. <https://doi.org/10.1007/s12639-019-01161-z>
- [14] MARTINS, W.M.D.O.; JUSTO, M.C.N.; CÁRDENAS, M.Q.; COHEN, S.C. Seasonality of parasitic helminths of *Leporinus macrocephalus* and their parasitism rates in farming systems in the Amazon. *Revista Brasileira de Parasitologia Veterinária* 26: 419-426, 2017. <https://doi.org/10.1590/s1984-29612017062>
- [15] BEEVI, M.R.; RADHAKRISHNAN, S. Community ecology of the metazoan parasites of freshwater fishes of Kerala. *Journal of parasitic diseases* 36: 184-196, 2012. <https://doi.org/10.1007/s12639-012-0101-8>
- [16] EIRAS, J.C. **Elementos de ictioparasitologia**. Fundação Eng. António de Almeida, Porto, Portugal. 1994.
- [17] SILVANO, R. A. M. **Peixes do Alto Rio Juruá (Amazonas, Brasil)**. EdUSP, 2001.
- [18] QUEIROZ, L. J. D.; VILARA, G. T.; OHARA, W. M.; PIRES, T. H. D. S.; ZUANON, J. A. S.; DÓRIA, C. R. **Peixes do Rio Madeira-Y-Cuyari Pirá-Ketá**. 2013.
- [19] EIRAS, J.C.; TAKEMOTO, R.M.; PAVANELLI, G.C. **Métodos de estudo e técnicas laboratoriais em parasitologia de peixes**. 2.ed. Eduem, Maringá. 2006.
- [20] SCHMIDT, G.D.; HUGGHINS, E.J. Acanthocephala of South American Fishes. Part I, Eoacanthocephala. *The Journal of Parasitology* 1: 829-835, 1973 DOI: 10.2307/3278417
- [21] PETTER, A.J. **Deux nouvelles espèces de Cucullanidae parasites de poissons en Guyane**. Bulletin du Muséum national d'Histoire naturelle, Paris. 1974.
- [22] MORAVEC, F. **Nematodes of freshwater fishes of the Neotropical Region**. Academia, Publishing House of the Academy of Sciences of the Czech Republic, 1998.
- [23] CAFFARA, M.; LOCKE, A.S.; GUSTINELLI, A.; MARCOGLIESE, D.J.; FIORAVANTI, M.L. Diferenciação morfológica e molecular de metacercárias e adultos de

*Clinostomum complanatum* e *Clinostomum marginatum* (Digenea: Clinostomidae). *Journal of Parasitology*, 97 (5), 884-891, 2011.

[24] REGO, A.A. Cestode parasites of neotropical teleost freshwater fishes. Metazoan parasites in the Neotropics: A systematic and ecological perspective 135-154, 2020.

[25] JONES, A.; BRAY, R.A.; GIBSON, D.I. **Keys to the Trematoda.**: CABI, Wallingford, UK. 2005.

[26] BUSH, A.O.; LAFFERTY, K.D.; LOTZ, J.M.; SHOSTAK, A.W. Parasitology meets ecology on its own terms: Margolis et al. revisited. **The Journal of parasitology** 1:575-583, 1997. DOI: 10.2307/3284227

[27] LUQUE, J.L.; POULIN, R. Metazoan parasite species richness in Neotropical fishes: hotspots and the geography of biodiversity. **Parasitology** 134:865-878, 2007. <https://doi.org/10.1017/S0031182007002272>

[28] BELLAY, S.; DE OLIVEIRA, E.F.; ALMEIDA-NETO, M.; ABDALLAH, V.D.; DE AZEVEDO, R.K.; TAKEMOTO, R.M.; LUQUE, J.L. The patterns of organisation and structure of interactions in a fish-parasite network of a Neotropical river. **International journal for parasitology** 45:549-557, 2015. <https://doi.org/10.1016/j.ijpara.2015.03.003>

[29] FROESE, R.; PAULY, D. (2017). FishBase 2017, version (march, 2017). World Wide Web electronic publication Home page at: <http://www.fishbase.org>.

[30] GUIDELLI, G.M.; ISAAC, A.; TAKEMOTO, R.M.; PAVANELLI, G.C. Endoparasite infracommunities of *Hemisorubim platyrhynchos* (Valenciennes, 1840)(Pisces: Pimelodidae) of the Baía River, upper Paraná River floodplain, Brazil: specific composition and ecological aspects. **Brazilian Journal of Biology** 63: 261-268, 2003. <https://doi.org/10.1590/S1519-69842003000200011>

[31] PINHEIRO, R.H.D.S.; FURTADO, A.P.; SANTOS, J.N.D.; GIESE, E.G. *Contraecaecum* larvae: morphological and morphometric retrospective analysis, biogeography and zoonotic risk in the amazon. **Revista Brasileira de Parasitologia Veterinária** 28:12-32, 2019. <http://dx.doi.org/10.1590/s1984-29612019002>.

[32] RUFFINO, M.L.; SILVA JUNIOR, U.L.; SOARES, E.C.; SILVA, C.O.; BARTHEM, R.B.; BATISTA, V.S.; ISAAC, V.J.; FONSECA, S.; PINTO, W. **Estatística Pesqueira do Amazonas e Pará.** IBAMA, Manaus. 2006.

[33] PAVANELLI, G. C.; REGO, A. A. *Megathylacus travassosi* sp. n. and *Nomimoscolex sudobim* Woodland, 1935 (Cestoda-Proteocephalidea) parasites of *Pseudoplatystoma*

- corruscans* (Agassiz, 1829)(Siluriformes-Pimelodidae) from the Itaipu reservoir and Paraná river, Paraná state, Brazil. **Memórias do Instituto Oswaldo Cruz**, v. 87, p. 191-195, 1992.
- [34] ZEHNDER, M.P.; DE CHAMBRIER, A. Morphological and molecular analyses of the genera *Peltidocotyle* Diesing 1850 and *Othinosclex* Woodland 1933, and a morphological study of *Woodlandiella* Freze, 1965 (Eucestoda, Proteocephalidea), parasites of South American siluriform fishes (Pimelodidae). **Systematic Parasitology**, 46:33-43, 2000.
- [35] REGO, A.A. Cestóides proteocefalídeos parasitas de *Pseudoplatystoma* (Pisces, Pimelodidae) da América do Sul. **Revista Brasileira de Zootecias**, 4(2), 2002.
- [36] SANTOS, K.R.; CATENACCI, L.S.; PESTELLI, M.M.; TAKAHIRA, R.K.; SILVA, R.J. First report of *Diphyllobothrium mansonii* (Cestoda, Diphyllbothridae) infecting *Cerdocyon thous* (Mammalia, Canidae) in Brazil. **Arquivo Brasileiro de Medicina Veterinária e Zootecnia** 56: 796-798, 2004. <http://dx.doi.org/10.1590/S0102-09352004000600016>
- [37] CAMPOS, C.M.; DE MORAES, J.R.E.; DE MORAES, F.R. Histopatologia do intestino de *Pseudoplatystoma fasciatum* (Osteichthyes, Pimelodidae) parasitados com cestodas proteocefalídeos e nematodas. **Boletim do Instituto de Pesca** 35:153-158, 2018.
- [38] RIBEIRO, T. S.; TAKEMOTO, R. M. Resposta inflamatória do pintado à infecção por *Nomimoscolex pertierra* (Eucestoda: Proteocephalidea). **Boletim do Instituto de Pesca**, v. 40, n. 1, p. 111-120, 2018.
- [39] ALVES, P.V.; DE CHAMBRIER, A.; LUQUE, J.L.; SCHOLZ, T. Towards a robust systematic baseline of Neotropical fish tapeworms (Cestoda: Proteocephalidae): amended diagnoses of two genera from the redbtail catfish, *Phractocephalus hemioliopus*. **Zootaxa**, 4370:363-380, 2018.  
DOI: <http://dx.doi.org/10.11646/zootaxa.4370.4.3>
- [40] CHAMBRIER, A.; KUCHTA, R.; SCHOLZ, T. Tapeworms (Cestoda: Proteocephalidea) of teleost fishes from the Amazon River in Peru: additional records as an evidence of unexplored species diversity. **Revue suisse de Zoologie** 122:149-163, 2020.  
<https://doi.org/10.5281/zenodo.14580>
- [41] DOGIEL, V.A.; PETRUSHEVSKI, G.K.; POLYANSKI, I. **Parasitology of fishes** (translated from Russian, 1967 by Kabata Z). England, TFH Ltd. 1970.
- [42] FROESE, R.; PAULY, D. (Eds.) (2020). FishBase. World Wide Web electronic publication. <http://www.fishbase.org>, version 01/2020.

- [43] LOCKE, S.A.; CAFFARA, M.; MARCOGLIESE, D.J.; FIORAVANTI, M.L. A large-scale molecular survey of *Clinostomum* (Digenea, Clinostomidae). **Zoologica Scripta** 44:203-217, 2015. <https://doi.org/10.1111/zsc.12096>
- [44] DE LEÓN, G.P.P.; GARCIA-VARELA, M.; PINACHO-PINACHO, C.D.; SERENO-URIBE, A.L.; POULIN, R. Species delimitation in trematodes using DNA sequences: Middle-American *Clinostomum* as a case study. **Parasitology** 143: 1773-1789, 2016. <https://doi.org/10.1017/S0031182016001517>
- [45] PÉREZ-PONCE DE LEÓN, G.; POULIN, R 2016. Taxonomic distribution of cryptic diversity among metazoans: not so homogeneous after all *Biology letters* 12: 20160371, 2016. <https://doi.org/10.1098/rsbl.2016.0371>
- [46] DE SOUZA, D.C.; CORREA, L.L.; TAVARES-DIAS, M. *Ithyoclinostomum dimorphum* Diesing, 1850 (Digenea, Clinostomidae) in *Hoplias malabaricus* (Erythrinidae) with the first report of infection of the eyes. **Helminthologia** 55: 343-349, 2018. <https://doi.org/10.2478/helm-2018-0028>
- [47] DIAS, M.L.G.; SANTOS, M.J.; SOUZA, G.T.R.; MACHADO, M.H.; PAVANELLI, G.C. Scanning electron microscopy of *Ithyoclinostomum dimorphum* (Trematoda: Clinostomidae), a parasite of *Ardea cocoi* (Aves: Ardeidae). **Parasitology research** 90:355-358, 2003. <https://doi.org/10.1007/s00436-003-0862-1>
- [48] BENIGNO, R.N.; KNOFF, M.; MATOS, E.R.; GOMES, D.C.; PINTO, R.M.; CLEMENTE, S.C. Morphological aspects of Clinostomidae metacercariae (Trematoda: Digenea) in *Hopleryttrinus unitaeniatus* and *Hoplias malabaricus* (Pisces: Erythrinidae) of the Neotropical region, Brazil. *Anais da Academia Brasileira de Ciências* 86: 733-744, 2014. <https://doi.org/10.1590/0001-3765201420130025>
- [49] KOHN, A.; MORAVEC, F.; COHEN, S.C.; CANZI, C.; TAKEMOTO, R.M.; FERNANDES, B.M. Helminths of freshwater fishes in the reservoir of the Hydroelectric Power Station of Itaipu, Paraná, Brazil. **Check List** 7:681-690, 2016. <http://dx.doi.org/10.15560/7.5.681>
- [50] SABAS, C.S.S.; BRASIL-SATO, M.C. Helminth fauna parasitizing *Pimelodus pohli* (Actinopterygii: Pimelodidae) from the upper São Francisco River, Brazil. **Revista Brasileira de Parasitologia Veterinária** 23:375-382, 2014. <https://doi.org/10.1590/1519-6984.15113>
- [51] ACOSTA, A.A.; QUEIROZ, J.; BRANDÃO, H.; SILVA, R.J.D. Helminth fauna of *Astyanax fasciatus* Cuvier, 1819, in two distinct sites of the Taquari River, São Paulo State,

Brazil. **Brazilian Journal of Biology** 75:242-250, 2015. <http://dx.doi.org/10.1590/1519-6984.15113>

[52] HERNÁNDEZ-MENA, D.I.; PINACHO-PINACHO, C.D.; GARCÍA-VARELA, . M.; MENDOZA-GARFIAS, B.; DE LEÓN, G.P.P. Description of two new species of allocreadiid trematodes (Digenea: Allocreadiidae) in middle American freshwater fishes using an integrative taxonomy approach. **Parasitology research** 118: 421-432, 2019. <https://doi.org/10.1007/s00436-018-6160-8>

[53] SANTOS, C.P.; GIBSON, D.I.; TAVARES, L.E.; LUQUE, J.L. Checklist of Acanthocephala associated with the fishes of Brazil. **Zootaxa** 1938: 1-22, 2008. <http://dx.doi.org/10.11646/zootaxa.1938.1.1>

[54] MARTINS, M.L.; DE MORAES, F.R.; FUJIMOTO, R.Y.; ONAKA, E.M.; QUINTANA, C.I.F. Prevalence and histopathology of *Neoechinorhynchus curemai* Noronha, 1973 (Acanthocephala: Neoechinorhynchidae) in *Prochilodus lineatus* Valenciennes, 1836 from Volta Grande Reservoir, MG, Brazil. **Brazilian Journal of Biology** 61:517-522, 2001. <https://doi.org/10.1590/S1519-69842001000300022>

[55] LIZAMA, M.D.L.A.P.; TAKEMOTO, R.M.; PAVANELLI, G.C. Influence of the seasonal and environmental patterns and host reproduction on the metazoan parasites of *Prochilodus lineatus*. **Brazilian Archives of Biology and Technology** 49:611-622, 2006. <https://doi.org/10.1590/S1516-89132006000500011>

[56] ARÉVALO, E.G.; MOREY, G.A.M.; DE OLIVEIRA MALTA, J.C. 2018. Fauna parasitária de *Prochilodus nigricans* (Prochilodontidae) de lagos de várzea da Amazônia Brasileira. **Biota Amazônia** 8:19-21, 2018. <http://dx.doi.org/10.18561/2179-5746/biotaamazonia.v8n1p19-21>

[57] LISITSYNA, O.; SCHOLZ, T.; KUCHTA, R. *Sharpilosentis peruviensis* ng, n. sp.(Acanthocephala: Diplosentidae) from freshwater catfishes (Siluriformes) in the Amazonia. **Systematic parasitology** 91:147-155, 2015. <https://doi.org/10.1007/s11230-015-9567-y>